Comparative wood and leaf anatomy of the Cecropiaceæ (Urticales)

K. Bonsen & B. J. H. ter Welle

Summary: The wood and leaf anatomy of the 6 genera of the Cecropiaceæ are described in detail. The anatomical data are compared with those of the allied Moraceæ and Urticaceæ. The relationship between habit, habitat, and anatomical characters is discussed, as well as the relationships within the family. Based on anatomical data the genus Poikilospermum should be included in the family Urticaceæ.

Résumé: Une analyse anatomique détaillée du bois secondaire et de la feuille des 6 genres constituant la famille des Cecropiaceæ est présentée. Les résultats ont été comparés à ceux obtenus pour des familles proches: Moraceæ et Urticaceæ. Les rapports entre les biotopes, les formes biologiques des plantes et les caractères anatomiques ont été considérés, ainsi que les affinités au sein de la famille. Les résultats anatomiques indiquent que le genre Poikilospermum doit être placé dans la famille des Urticaceæ.

Karel Bonsen & Ben J. H. ter Welle, Department for Systematic Botany, Heidelberglaan 2, 3584 CS Utrecht, The Netherlands.

INTRODUCTION

Recent taxonomic studies of the Moraceæ, especially those from tropical America and Africa, have been carried out by C. C. Berg, Institute for Systematic Botany (Utrecht), while anatomical studies of this family were conducted by S. M. C. Topper (wood) and A. Kloos (leaf) also at Utrecht. In 1978, Berg separated the Cecropiaceæ from the Moraceæ. A detailed anatomical study of this new family to complete the leaf and wood anatomical survey, therefore seemed desirable.

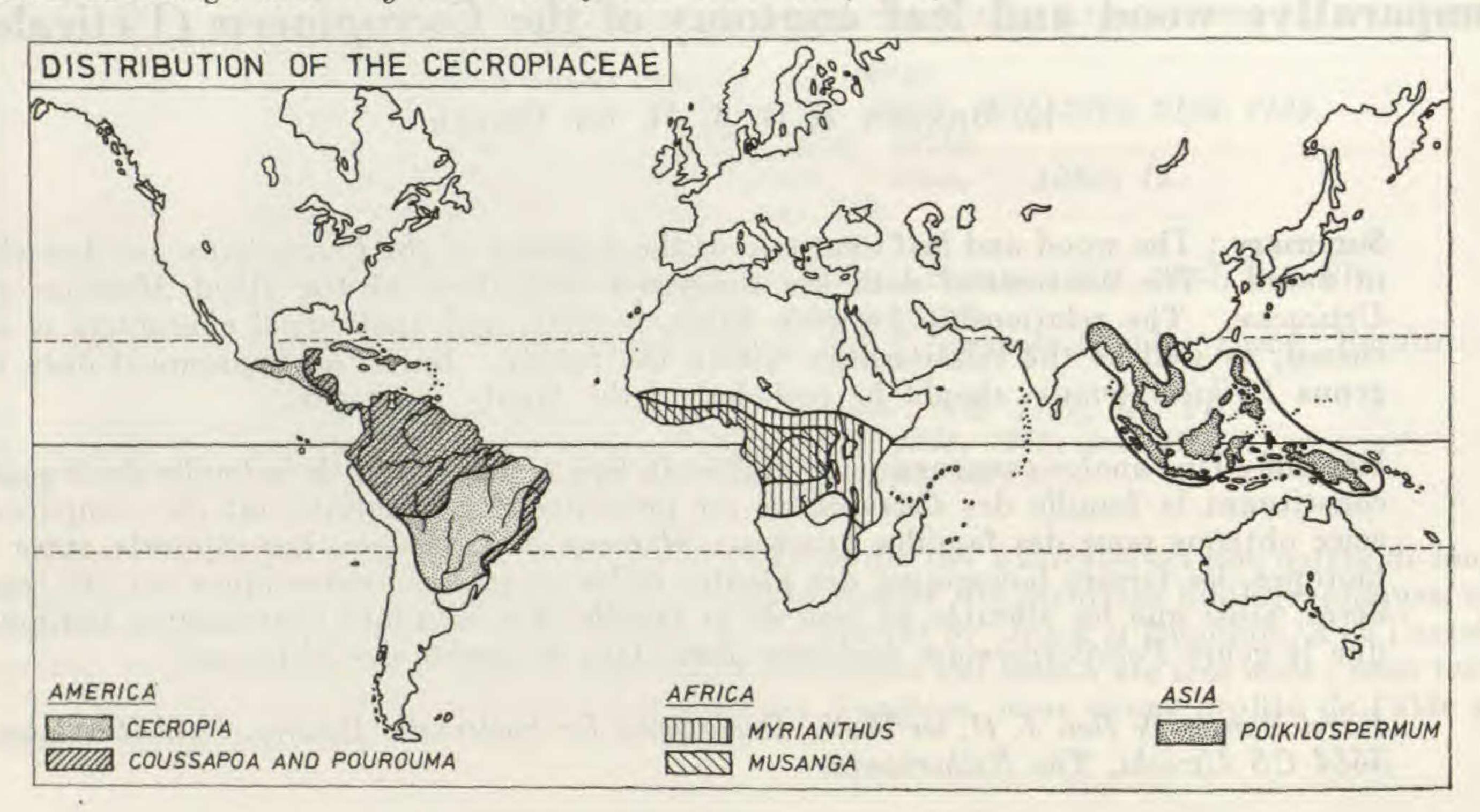
The Cecropiaceæ constitute a pantropical family of about 200 species in 6 genera. Among the genera we find hemi-epiphytic scramblers, shrubs or trees with aerial roots, sometimes lianas, and small to tall trees, often with stilt-roots. Most species of Coussapoa and Poikilospermum are hemi-epiphytes, while the species of Cecropia and Musanga are commonly pioneer plants, and most species of Pourouma and Myrianthus are small or medium-sized trees of the lower stories of more or less open and disturbed forests.

In Engler's system of the Moraceæ (1889) the neotropical genera Cecropia, Coussapoa, and Pourouma, together with the African genera Musanga and Myrianthus, and the Asiatic genus Poikilospermum (= Conocephalus), constituted the subfamily Conocephaloideæ. Chew Wee-Lek (1963) suggested to transfer the small-seeded genera Cecropia, Coussapoa, Musanga, and Poikilospermum to the Urticaceæ, but retained the big-seeded genera Myrianthus and Pourouma in the Moraceæ. Corner (1962) transferred the whole subfamily

Conocephaloideæ to the Urticaceæ mainly on the basis of similarities in the characters of the ovary: a single stigma and a basally attached orthotropous ovule.

The genus Poikilospermum has been monographed by Chew Wee-Lek (1963), the

genera Musanga and Myrianthus by DE Ruiter (1976).



Anatomical studies of the subfamily Conocephaloideæ (= Cecropiaceæ) were made by Renner (1907), and of Cecropia by Richter (1898). Renner noticed the isolated position of the genus Conocephalus (= Poikilospermum) based on the structure of the stomata and glands, and the presence of cystoliths. He pointed out the remarkable anatomical similarity between the American and African genera and showed that these genera fall into two groups on the basis of their adaxial glands; viz. Musanga, Myrianthus and Pourouma on the one hand and Cecropia and Coussapoa on the other. More anatomical data have been given by Solereder (1899, 1908), and Metcalfe & Chalk (1950).

Finally, Berg (1978) created the Cecropiaceæ also using anatomical data as far as

known to him from literature.

MATERIALS AND METHODS

The majority of the wood samples for this study was taken from the Utrecht wood collection, while the leaves were taken from vouchers deposited in the herbarium in Utrecht. Herbarium vouchers of all specimens, identified by C. C. Berg and E. G. B. Kieft (Pourouma) are located in Utrecht. Information on collector's numbers and wood collection accession numbers (abbreviations according to Stern, 1978), locality and diameter of the samples, is given in the tables 1 and 2. This study is based on 84 wood specimens representing 6 genera and 45 species, apart from 45 leaf specimens representing 6 genera and 34 species. Sections and macerations were prepared according to standard techniques, and embedded in Canada balsam and in glycerin respectively. The wood anatomical terminology used is that proposed by the Committee on Nomenclature of the I.A.W.A. (1964).

SPECIES	LEAF	N HERBARIUM	Corrector	COLLECTION
	COLLECTIO	NILERBARIUM	COLLECTOR	LOCALITY
Cecropia latiloba Miq.	B-404	332521 B	Prance & Berg P 17586	Brasil, Amazonas
C. obtusa Trec.	B-405	378826 B	Prévost 732	Fr. Guiana
C. sciadophylla Mart.	B-406	258202 B	Roberts L.B.B. 12791	Surinam
C. surinamensis Miq.	B-407	253593 B	L.B.B. 12739	Surinam
C. sp.	B-459		Hortus Baarn 2/7, cult.	Netherlands (Cult.)
Coussapoa angustifolia Aubl.	B-420	211911 B	van Donselaar 2569	Surinam
C. asperifolia Trec.	B-436	362253 B	Oldeman B 2538	Fr. Guiana
C. latifolia Aubl.	B-416	81393 B	Mennega 271	Surinam
))	B-417	332696 B	Prance & al. 58783	Brasil, Para
))	B-437	332533 B	Lieras & Monteiro P 19661	Brasil, Lara
))	B-438	384674 B	Heyde & Lindeman 78	Surinam
C. microcarpa (Schott) Rizzini	B-421	270318 B	Lindeman & al. 5682	Brasil, Parana
C. microcephala Trec.	B-439	390327 B	Maas 2478	Guyana
C. nitida Miq.	B-422	332608 B	Berg & al. P 17592	Brasil, Amazonas
))	B-440	393714 B	Huashikat 1585	Peru Peru
C. orthoneura Standl.	B-418	362170 B	Revilla 1847	Peru
))	B-419	277078 B	Prance & al. 14029	Brasil, Amazonas
))	B-441	393716 B	Liesner 8750	Venezuela
C. ovalifolia Trec.	B-442	393720 B	Diaz & Osores 618	Peru
C. panamensis Pitt.	B-443	370065 B	Neill L984	
C. parvicops Standl.	B-444	370061 B	Hammel 2176	Nicaragua Panama
C. trinervia Mildbr.	B-445	393718 B	Rosa & Vilar 2874	Brasil
C. villosa P. & E.	B-446	393717 B	Pires & Santos s.n.	Brasil
Musanga cecropioides R. Br.	B-408	257909 B	Versteegh & al. 52	
))	B-450	159447 B	Mann s.n.	W. trop. Africa
Myrianthus arboreus Pal. Beauv.	B-411	41498 B	Deistel 428	Cameroon
M. libericus Rendle	B-409	257646 B	Versteegh & al. 24	Ivory Coast
M. serratus (Trec.) B. & H.	B-410	202186 B	Mann 2273	W. trop. Africa
Poikilospermum abnorme Chew	B-415	11223 B	Eyma 3536	Indonesia, Celebes
P. amboinense Zipp. ex Miq.	B-412	34062 A	Docters van Leeuwen	Indonesia, NW. Guinea
))	B-413	34111 A	Docters van Leeuwen	Indonesia, NW. Guinea
P. inæquale Chew	B-403		Jacobs 9575	Papua, NW. Guinea
P. subtrinervium (Miq.) Chew	B-414	33780 A	Mandi 25	Indonesia, Borneo
Pourouma acuminata Mart. ex Miq.	B-580		Jobert 682	Brasil, Potomayo
P. albistipulata	B-396		Steyermark & al. 103026	Venezuela
P. cecropiifolia Mart.	B-579		Krukoff 5327	Brasil, Acre
P. cuspidata Warb.	B-397	38698 A	Krukoff 8373	Brasil, Amazonas
P. feruginea Standl.	B-398		Coêlho & al. 339	Brasil, Amazonas
P. guianensis Aubl.	B-399	355615 B	Berg & al. BG 772	Brasil, Para
P. lævis Benth.	B-400	673930 B	Boschwezen 4010	Surinam
P. minor R. Ben.	B-014	358752 B	Schunke 8487	Peru, Amazonas
P. mollis Trec.	B-581		van Donselaar 1681	Surinam
P. velutina Miq.	B-447	361783 B	Prance 21011	Brasil, Amazonas
P, sp .	B-448		Berg & Nee 355	Panama, Canal Zone
P. sp.	B-449		Herbario Adolfo Tonduz 12930	

Table 2: Wood specimens of the Cecropiaceæ studied.

SPECIES	XYLARIUM	CGLLECTOR	COLLECTION	MINIMUM DIAMETER (cm)
	U 25025	Cuatrecasas 15260	Colombia, Choco	10
Cecropia burriada Cuatr.		Prance & Berg P 19817	Brasil, Amazonas	9
C. concolor Willd.	U 20959	Prance & Berg P 18526	Brasil, Amazonas	9
C. ficifolia Snethl.	U 20919		Colombia, Choco	10
C. garcia Standl.	U 27511	Cuatrecasus 16467	Equador	2
C. monostachya C. Berg	U 23612	Berg, Maas & ter Welle 416	Guyana	10
C. obtusa Trec.	U 21569	Smith 2164	Cuba	
C. peltata L.	U 25719	Süss 17	Guyana	
C. riparia Warb.	U 21655	Smith 3426	Surinam	10
C. sciadophylla Mart.	U 179	Stahel 179		9
))	U 2146	B.B.S. V16	Surinam	10
))	U 2296	Ellenberg	Peru	6
))	U 2379	Lindeman 3617	Surinam	6
))	U 5124	BAFOG~39M	Fr. Guiana	9
))	U 7581	Krukoff 6237	Brasil, Amazonas	15
))	U 17503	Maguire & al. 55577	Surinam	7
))	U 19622	Krukoff 4789	Brasil, Amazonas	9
))	U 19929	Krukoff 5389	Brasil, Acre	2
))	U 23290	Heyde 710	Surinam	
C. telealba Guatr.	U 27512	Cuatrecasas 18353	Colombia, Choco	14
Comment Calin And I	U 4470	Lindeman 6545	Surinam	9
Coussapoa angustifolia Aubl.	U 3917	Lindeman 5709	Surinam	25
C. asperifolia Trec.	Dw 5596	Littuemun 9100	Brasil	10
O L. L. Al-L. S. Dane	U 25671	v. Rooden, ter Welle & Topper 700	Colombia, Choco	
C. batavorum Akk. & Berg	U 25192	Cuatrecasas 14284	Colombia, Choco	6
C. contorta Cuatr.		Berg & Akkermans 1142	Equador	
C. hololeuca Miq.	U 27072	Lindoman & Houde 78	Surinam	15
C. latifolia Aubl.	U 22728	Lindeman & Heyde 78	Equador	6
C. trinervia Spruce	U 27032	Berg & Akkermans 1038	Brasil, Amazonas	8
C. villosa P. & E.	U 20911	Prance & Berg P. 18455	Drash, Amazonas	

TABLE 2 (Contd.).

SPECIES	XYLARIUM	COLLECTOR	COLLECTION	MINIMUM DIAMETER (CD
Musanga cecropioides R. Br.	U 6445	ex RBHw	Cameroon	19
))	U 24230	Louis 13501	Congo	30
))	U 24231	Donis 435	Congo	
))	U 24386	Antoine 283	Congo	30
))	U 24489	Timorno 200	Angola	50
))	U 24613	ex MAD-SJRw 15253	Liberia	90
	U 24614	ex MAD-SJRw 15799	Liberia	20
"	U 24622	ex MADw 36807		20
3)	Dw 5609	Antoine s.n.	Ivory Coast	20
M. leo-ereræ Haum.	U 24232	Machine Machin	Zaïre	20
in. teo-ereræ Haum.	0 24232	Bauxin 1021	Rwanda	15
Myrianthus arboreus Pal. Beauv.	U 20313	de Briey 49	Zaire	
))	U 24385	R 478-80	Tanganyika	15
»	U 24446	Leeuwenberg 9656	Cameroon	7
))	U 24619	ex MADw 36828	Ivory Coast	10
))	U 24632	ex MADw 32716	Congo	
M. holstii Engl.	U 15509	ex RBHw 1542	East Africa	8
))	U 15525	ex RBHw 1577	East Africa	8
))	U 24234	Bouxin 1291	Rwanda	
M. libericus Rendle	U 24235	Cooper 285	Liberia	
))	U 24615	ex MAD-SJRw 13778	Liberia	
))	U 24617	ex MAD-SJRw 15198	Liberia	
))	U 24621	ex MADw 36781	Ivory Coast	15
M. serratus (Trec.) B. & H.	U 24618	ex MAD-SJRw 15110	Liberia	4
Poikilospermum amboinense Zipp. ex Miq.	U 26781	Lam 647	Indonesia, NW Guinea	1,5
P. inæquale Chew	U 26780	Docters v. Leeuwen 9671	Indonesia, NW Guinea	1
P. naucleiflorum Euse		1507/i22-H 1868-274	Theorem, The Countries	2
P. suaveolens (Bl.) Merr.	U 27070	Jacobs 8502	Indonesia, Sumatra	3,5
))	Pw/U 27516	de Vogel 4516	Indonesia, N. Moluccas	9,0
P. sp.	Lw/U 27517	Jacobs 9575	Papua NW Guinea	9
P. sp.		Koorders 35779 B	Indonesia, Java	1,5

TABLE 2 (Contd.)

Species	XYLARIUM	COLLECTOR	COLLECTION LOCALITY	MINIMUM DIAMETER (cm)
	TTACAGC	Krukoff 8427	Brasil, Amazonas	10
Pourouma acuminata Mart. ex Miq.	U 16196 U 13744	Hatschbach & Lindeman 13537	Brasil, Parana	15
P. acutiflora Trec.	Dw 5640	Huistinate & Lincollium 1999	Brasil	10
P. apiculata Spruce	U 25405	Cuatrecasas 15071	Colombia	8
P. aspera Trec.		Krukoff 5109	Brasil, Amazonas	18
P. cecropiifolia Mart.	U 19763 U 19903	Krukoff 5327	Brasil, Acre	4
"	U 24925	Cuatrecasas	Colombia, Choco	5
P. chocoana Standl.	Dw 2225	M 2214		10
P. digitata Trec.	U 17970	Oldenburger, Norde & Schulz 1406	Surinam	11
"	U 11140	Florschütz & Maas 3132	Surinam	4
P. guianensis Aubl.		Krukoff 1297	Brasil, Amazonas	
))	U 19274	Prance & Berg P 18250	Brasil, Amazonas	20
))	U 20956	Smith 2845	Guyana	
))	U 21586	de Bruyn 1546	Colombia	12
P. hirsutipetiolata Mildbr.	U 14494	Cuatrecasas 14881	Colombia, Choco	2
P. hispida Standl.	U 25420	Maguire & al. 55576	Surinam	17
P. lævis Benth.	U 17502	Lanjouw & Lindeman 432	Surinam	25
P. maroniensis R. Ben.	U 1233 U 21508	Smith 2731	Guyana	
n	U 1208	Lanjouse & Lindeman 399	Surinam	15
P. melinonii R. Ben.	U 20897	Prance & Berg P 18136	Brasil, Amazonas	25
)) 77 • • • • • • • • • • • • • • • • • •	U 8669	Ellenberg 2297	Peru	
P. mollis Trec.	U 8155	Krukoff 7073	Brasil, Amazonas	10
P. ovata Trec.	Dw 5636	USw 6268	Brasil	10
P. substrigosa Mildbr.	U 19886	Krukoff 5309	Brasil, Acre	13
P. triloba Trec.	U 27206	Mexia s.n.	Peru	
P. sp.	U 27513	Dusén 17345	Brasil, Parana	1,5
P. sp.	0 2/313	Dusch 11010		
URTICACEÆ				
** ** *** ***	TT 94000	Schunke 4977	Peru	2
Boehmeria pavonii Wedd.	U 21090	Chambers 2686	Dominica	4
B. ramiflora Jacq.	U 15446	USw 6019	Jamaica	2
Gyrotænia microcarpa F. & R.	U 8357	Schunke 4008	Peru	1,5
Myriocarpa stipitata Benth.	U 21084		Hawaii	4
Touchardia latifolia Gan.	The state of the s	Stern & Herbst 518 Mathias & Taylor 5343	Peru	1
Urera elata (Sw.) Griseb.	U 27194	Mathias & Taylor 5343 Schlieben 1721 ex RBHw	East Africa	3
U. hypselodendron (Horhst.) Wedd.	U 15936		Zimbabwe	3
))	U 27404	Berg		

The quantitative wood data were measured as follows: vessel diameters were measured in tangential direction; averages are based on 25 measurements. The vessel frequency is based on 25 counts of areas of 1 sq. mm. In the descriptions, average, minimum and maximum values are given for both characters. The percentage of solitary vessels was calculated after examining an area showing at least 100 pores. Clusters and multiples were regarded as 2, 3, 4, etc. vessels, depending on the number of vessels per group. For the intervascular pits the minimum and maximum sizes are given. Vessel member length, fibre length and parenchyma length (including both strands and fusiform cells) are based on 25 measurements per sample. Averages, minimum and maximum sizes are given. Additionally, the averages were used to calculate the ratio of fibre length/vessel element length, in the descriptions referred to as F/V ratio. For the fibres, maximum wall thickness, maximum lumen diameter and the average ratio of lumen diameter/ wall thickness (= twice maximum wall thickness), in the descriptions referred to as L/W ratio, are given. Multiseriate ray height is presented in micrometers (µm), ray width in number of cells. The data concern the averages of the 25 highest rays as observed in each section. Uniseriate ray height is based on 25 measurements per section. The percentage of uniseriate rays taken from the total number of uniseriate and multiseriate rays, and the percentage of vertically compound multiseriate rays is reported. The number of rays per mm is the average of 25 counts. The epidermal cells of the leaves as seen in the greater part of the samples were hardly measurable (e.g. covered by arachnoid hairs). In the descriptions the average dimensions of the epidermal cells are called tall (35-45 μm), small 25-35 μm), or very small (less than 25 μm). The shape of the periclinal walls is only mentioned if it is not straight or faintly sinusoid. "Intermediate layers" are layers of palisade-like tissue, consisting of conjugated spongy cells, lying between the palisade tissue and the spongy tissue. The midrib vascular system was studied at one third the lamina length from the leaf base. The petiole vascular system was studied at one half the length of the petiole.

GENERIC DESCRIPTIONS

I. SECUNDARY XYLEM

1. Cecropia Loefling - Pl. 2, 1.

Studied: 10 species, 19 specimens.

A genus of probably 70-80 species in tropical America, forming small to tall trees often with stilt-roots.

Growth rings faint or absent. Vessels diffuse, round to oval, solitary (50-82 %) and in radial multiples and irregular clusters of 2-3 (5), 1-3 (0-9) per sq. mm, diameter 155-300 (150-350) μ m, vessel member length 500-690 (400-800) μ m. Perforations simple. Intervascular pits alternate, round or polygonal, 12-18 μ m. Thin-walled tyloses present or absent.

Fibres non septate, diameter 26-50 μ m, walls 2-4 (6) μ m, L/W ratio 1.6-10, gelatinous fibres scarce. Pits simple, mainly on the radial walls. Length 1100-2100 (975-2400) μ m,

F/V ratio 2.2-3.5.

Rays heterogeneous, uniseriate (3-25 %) and multiseriate, 3-7 (2-10) per mm, sheath cells scarcely present or absent. Uniseriate rays mainly composed of square to upright cells, ray height 300-700 (200-950) µm. Multiseriate rays composed of upright and procumbent cells, vertically compound 0-27 %, 1000-2200 (500-3250) µm high, 2-6 cells in width, uniseriate parts 1-4 (0-16) cells, sometimes containing rhombic crystals.

Parenchyma scarce. Paratracheal parenchyma vasicentric to aliform, sometimes confluent. Strands 4-6 (8) cells, length 570-770 (510-870) µm, sometimes containing rhombic crystals. Apotracheal parenchyma terminal, sometimes consisting of 2 narrow, concentric bands, present or absent.

Specific gravity: 0.25-0.55.

Note: Radial latex tubes were observed in a few samples of C. sciadophylla Mart. and in C. monostachya C. C. Berg.

2. Coussapoa Aublet — Pl. 2, 3.

Studied: 8 species 9 specimens.

A genus of 49 species in tropical America. Usually hemi-epiphytic shrubs or trees with aerial roots or with stilt-roots if terrestrial.

Growth rings faint or absent. Vessels diffuse, round to oval, solitary (25-83 %) and in radial multiples and irregular clusters of 2-8 (21), 1-6 (0-11) per sq. mm, diameter 220-300 (200-340) µm. Vessel member length 475-600 (400-725) µm. Perforations simple. Intervascular pits alternate, round or polygonal, 10-15 µm. Thin-walled tyloses usually present.

Fibres non septate, diameter 18-25 μm, walls 2-3.5 μm, L/W ratio 2-5, gelatinous fibres usually present. Pits simple, mainly on the radial walls. Length 1100-1800

(875-2175) μm, F/V ratio 2.5-3.7.

Rays heterogeneous, uniseriate (21-35 %) and multiseriate, 4-7 (3-9) per mm, sheath cells present or absent. Uniseriate rays mainly composed of square to upright cells, ray height 300-500 (200-980) µm. Multiseriate rays composed of upright and procumbent cells, vertically compound 0-10 %, 700-1100 (450-1600) µm high, 3-6 cells in width, uniseriate parts 1-2 (0-8) cells.

Parenchyma paratracheal, banded, irregular, wavy, 1-2 (0-3) per mm, 5-9 (3-15) cells in width. Strands 5-8 (14) cells, length 600-710 (530-870) µm, containing some to many

rhombic crystals.

Specific gravity: 0.50-0.75.

Note: Radial latex tubes observed in C. latifolia Aublet.

3. Musanga R. Brown — Pl. 2, 2.

Studied: 2 species, 10 specimens.

A genus of 2 species in tropical Africa forming trees with stilt-roots.

Growth rings faint or absent. Vessels diffuse, round to oval, solitary (50-93 %) and in radial multiples and irregular clusters of 2-3 (4), 1-2 (0-4) per sq. mm, diameter 210-310 (190-350) µm, vessel member length 450-605 (350-725) µm. Perforations simple. Intervascular pits alternate, round to polygonal, 12-15 (18) µm. Thin-walled tyloses usually present.

Fibres non septate, diameter 34-54 μ m, walls 1-3.5 μ m, L/W ratio 5-over 25, gelatinous fibres present or absent. Pits simple, mainly on the radial walls. Length 1150-1950 (975-2400) μ m, F/V ratio 2.1-3.2.

Rays heterogeneous, uniseriate (5-11 (23) %) and multiseriate, 3-5 (2-7) per mm, sheath cells absent. Uniseriate rays mainly composed of square to upright cells, ray height 270-470 (200-720) μm. Multiseriate rays composed of upright and procumbent cells, vertically compound 0-60 %, 750-1100 (500-1850) μm high, 2-4 cells in width, uniseriate parts 1-3 (0-9) cells, often containing rhombic crystals.

Parenchyma scarce. Paratracheal parenchyma vasicentric to aliform, strands 4-5 (12) cells, length 625-775 (550-950) µm, often containing rhombic crystals. Apotracheal

parenchyma terminal, present or absent.

Specific gravity: 0.12-0.40.

4. Myrianthus Pal. Beauv. — Pl. 2, 4.

Studied: 4 species, 13 specimens.

A genus or 7 species in tropical Africa. Usually medium-sized trees or shrubs with stilt-roots or sometimes lianas.

Growth rings faint or absent. Vessels diffuse, round to oval, solitary (30-85 %) and in radial multiples and irregular clusters of 2-3 (5), 2-6 (1-11) per sq. mm, diameter 120-250 (110-275) μ m, vessel member length 400-550 (300-675) μ m. Perforations simple. Intervascular pits alternate, round or polygonal, 10-15 μ m. Thin-walled tyloses present or absent.

Fibres non septate, diameter 18-29 μm , walls 2.2-5 μm , L/W ratio 2-5, gelatinous fibres present or absent. Pits simple mainly on the radial walls. Length 1100-1850

(950-2125) µm, F/V ratio 2.5-4.0.

Rays heterogeneous, uniscriate (12-50 %) and multiseriate, 4-8 (2-10) per mm, sheath cells present or absent. Uniscriate rays mainly composed of square to upright cells, ray height 350-850 (250-1450) µm, multiseriate rays composed of upright and procumbent cells, vertically compound 0-10 %, 1000-1750 (700-2500) µm, 4-10 cells in width, uniscriate parts 1-4 (0-15) cells, sometimes containing rhombic crystals.

Parenchyma banded, irregular, wavy, 1-2 (3) par mm, 4-12 (3-16) cells in width. Strands 4-5 cells, length 420-670 (400-900) p.m, containing some to many rhombic crystals.

Specific gravity 0.45-0.60.

5. Poikilospermum Zipp. ex Miquel — Pl. 3, 1.

Studied: 4 species, 7 specimens.

A genus of 20 species in tropical Asia. Hemi-epiphytic scramblers with aerial roots.

Growth rings absent. Vessels diffuse, round to oval, solitary (55-88 %) and in radial multiples and irregular clusters of 2-3 (9), 5-9 (3-12) per sq. mm, diameter 260-310 (180-400) μ m, vessel member length 325-385 (250-475) μ m. Perforations simple. Intervascular pits alternate, round or polygonal, 15-20 μ m. Thin-walled tyloses present or absent.

Fibres showing dimorphism: short fibres, length 545-865 (400-1100) μ m, non septate, diameter 22-26 μ m, walls 2.5-3.5 μ m, L/W ratio, 3.5-5, gelatinous fibres scarce. Pits simple, on radial and tangential walls. F/V ratio 1.6-2.4. Very long fibres, length 4000-5000 μ m.

Rays heterogeneous, multiseriate, partly unlignified, composed of upright and procumbent cells, sometimes vertically compound to vertical rows, 1540-2875 (700-4700) µm

high, 4-9 cells in width, uniseriate parts absent, 1-3 per mm.

Parenchyma: paratracheal parenchyma vasicentric; apotracheal parenchyma in irregular unlignified concentric bands. Strands 370-450 (275-625) µm, containing druses and often rhombic crystals.

Specific gravity unknown.

Note: Juvenile parts differ in many characters from the foregoing generic description:

Vessels 9-22 (3-33) per sq. mm, diameter 100-150 (85-200) µm, vessel member length

295-345 (200-450) µm.

Fibres, diameter 14-16 μ m, walls 3-4 μ m, L/W ratio 2-3. Very long fibres absent. Rays heterogeneous or homogeneous, unlignified parts scarce or absent, 4 (2-7) per mm. Unlignified apotracheal parenchyma absent or scarce.

6. Pourouma Aublet

Studied: 17 species, 26 specimens.

A genus of probably more than 50 species in tropical America. Small or medium-sized trees, often with stilt-roots.

Growth rings faint or absent. Vessels diffuse, round to oval, solitary (55-95 %) and in radial multiples and irregular clusters of 2-3 (4), 1-6 (0-8) per sq. mm, diameter 125-255 (110-305) μ m, vessel member length 475-850 (350-930) μ m. Perforations simple. Intervascular pits alternate, round or polygonal, 10-20 μ m. Thin-walled tyloses present or absent.

Fibres non septate, diameter 18-36 μm, walls 1.2-4 μm, L/W ratio 3-9, gelatinous fibres present or absent. Pits simple, mainly on the radial walls. Length 940-1725 (775-2070) μm, F/V ratio 1.6-2.7.

Rays heterogeneous, uniseriate (5-53 %) and multiseriate, 4-9 (3-12) per mm, sheath cells present or absent. Uniseriate rays mainly composed of square to upright cells, ray height 300-750 (200-1000) µm, multiseriate rays composed of upright and procumbent cells, vertically compound 8-23 (45 %), 450-1260 (1800) µm high, 2-5 cells in width, uniseriate parts 1-6 (0-16) cells, sometimes containing rhombic crystals.

Parenchyma paratracheal variable, vasicentric and aliform to confluent and even banded. Bands irregular, wavy, 2-3 per mm, 5-6 (4-8) cells in width. Strands 5-6 (7) cells, length 560-950 (450-1000) µm, often containing rhombic crystals. Apotracheal parenchyma terminal, sometimes consisting of 2 narrow, concentric bands, present or absent.

Specific gravity 0.40-0.75.

Note: Radial latex tubes observed ino ne sample of P. melinonii R. Ben. The multiseriate rays of P. triloba Trec. are much higher than those in the foregoing description: 2280 (1500-3500) μm.

Discussion on wood anatomical characters

The family Cecropiaceæ can be divided into two groups: the genus Poikilospermum and the genera Cecropia, Musanga, Coussapoa, Myrianthus and Pourouma. The latter group can be sub-divided into the Cecropia-Musanga group and the Coussapoa-Myrianthus group, with Pourouma overlapping both groups. These divisions can be made with regard to many wood characteristics, for example (see also table 6): the average number of vessels per sq. mm, 1-3 in Cecropia and Musanga, 1-6 in Coussapoa, Myrianthus and Pourouma, 5-9 in Poikilospermum; the average vessel member length, 325-385 µm in Poikilospermum and 400-850 µm in the other genera; the location of the fibre pits which are on both radial and tangential walls in Poikilospermum and on radial walls only in the other genera; the average fibre length, 545-865 µm for the short fibres and 4000-5000 μm for the long fibres in Poikilospermum varies between 940 and 2100 μm in the other genera; the rays with exception of Poikilospermum consist of 2 types, uniseriates (5-53 %) and multiseriates; the number of rays per mm, 1-3 in Poikilospermum and 3-9 in the other genera; Poikilospermum is the only genus with unlignified ray parts and unlignified apotracheal parenchyma; the average parenchyma strand length, 370-450 µm in Poikilospermum and 420-950 µm in the other genera; the crystal type, druses and rhombic crystals in Poikilospermum and rhombic crystals only in the other genera.

The genus Poikilospermum consists of hemi-epiphytic scramblers. The unlignified parenchyma and the reticulate parenchyma pattern in this genus might be related to the climbing habit (TER Welle & Koek-Noorman, 1981). Because of the facts that there are non-climbing members of the Urticaceæ which also have unlignified parts and that all (climbing) members of the Moraceæ lack this phenomenon, this character supports the taxonomic separation of Poikilospermum from the other genera. Species of the genus Coussapoa are usually hemi-epiphytic. The only indication of a correlation between this habit and the characters found might be the number of vessels per multiple, which is also observed in the hemi-epiphytic genus Poikilospermum. Pourouma and Myrianthus are small to medium-sized trees, Cecropia and Musanga are pioneer plants (Berg, 1978, 1981). These two groups of genera show similarities in characters. The montane species like Myrianthus holstii Engl. and Musanga leo-erreræ Haum. show a smaller pore and fibre diameter, and shorter vessel members and fibres than the lowland species like Myrianthus arboreus Pal. Beauv. and Musanga cecropioides R. Br. (cf. van den Oever, Baas & Zandee,

1981).

Considering the range of diversity of the wood characters of the Moraceæ, the Cecropiaceæ could be placed in the Moraceæ on the basis of their wood structure (Mennega, pers. comm. in Berg, 1978; Topper, pers. comm.). This may be true concerning the genera Cecropia, Coussapoa, Musanga, Myrianthus and Pourouma, particularly because of the presence of latex tubes in some samples. These 5 genera are characterized by diffusely distributed vessels, solitary (25-95 %) and in radial multiples and irregular clusters of 2-8 (21), 1-6 (0-11) per sq. mm, diameter 120-310 (110-350) μm, vessel member length 400-850 (300-930) μm.

Perforations simple. Intervascular pits alternate, round or polygonal, 10-18 μm. Fibres non septate, diameter 18-54 μm, walls 1-5 (6) μm, L/W ratio 1.6-over 25. Pits simple, on radial walls. Length 940-2100 (775-2400) μm, F/V ratio 1.6-4.0. Rays heterogeneous, uniseriate (0-53 %) and multiseriate, 3-9 (2-12) per mm. Uniseriate rays mainly composed of square to upright cells, ray height 270-850 (200-1450) μm. Multiseriate rays composed of upright and procumbent cells, vertically compound 0-60 %, 450-2200 (350-3250) μm high, 2-10 cells in width, uniseriate parts 1-6 (16) cells. Paratracheal parenchyma vasicentric, aliform, confluent or banded; bands irregular, wavy, 1-3 per mm, 4-12 (3-16) cells in width. Strands 420-950 (400-1000) μm, 3-8 (4-11) cells, often containing rhombic crystals. Specific gravity 0.12-0.75.

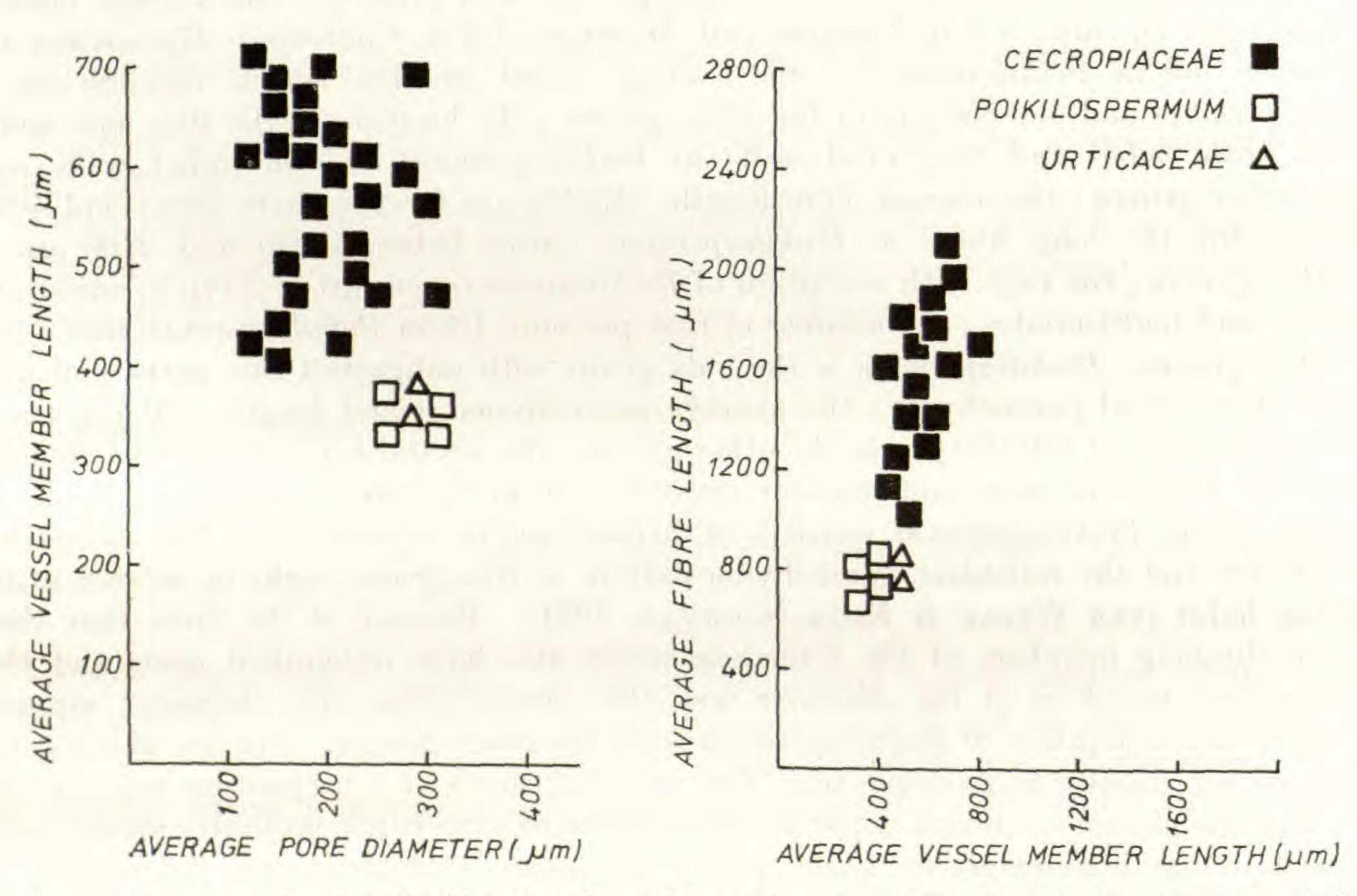


Fig. 1. — Vessel member length, fibre length, and pore diameter of the Cecropiaceæ, the genus Poikilospermum and some representatives of the Urticaceæ.

Based on data from literature (Metcalfe & Chalk, 1950) and on our own observation the wood of *Poikilospermum* seems to be rather urticaceous. Its vessel diameter, F/V-ratio (see fig. 1), location of the fibre pits, ray type, number of rays per mm, the presence of unlignified parts, and the crystal type are in agreement with features observed in species of the genera *Boehmeria*, *Gyrotænia*, *Myriocarpa*, *Touchardia*, *Urera* and *Urtica*. The presence of the 4000-5000 mm long fibres which occur in *Poikilospermum* could not be demonstrated in the *Urticaceæ*. Some of the species of the tribes *Urereæ* and *Boehmerieæ* have unlignified parenchyma in the wood, part of these species are climbers or lianas (e.g. *Urera hypselodendron*, Pl. 3,2), the others are non-climbing shrubs or herbs (e.g.

Gyrotænia, Laportea, Myriocarpa, Touchardia, Urtica dioica, Urera elata). Table 3 shows that the wood of Poikilospermum closely resembles the wood of the climbing representatives of the tribe Urereæ.

Table 3: Comparison of relevant characters of the genus Poikilospermum and some urticaceous taxa.

	U. hyps. (A)	Poik. (A)	Poik. (J)	U. hyps. (J)	Boehmeria	Urticaceæ
Vessel diameter (µm)	290-350	260-310	100-150	120-180	90-120	110-140
Vessel member length (µm)	375	325-385	295-345		330-475	445-520
Vessels per sq. mm	4-10	5-9	9-22	8-12	17-23	8-16
Fibre length (µm)	630-810	550-870	655-810		650-730	725-920
Fibre pit location		R, T	R, T	R, T	R, T	R, T
Rays per mm	R, T 1-3	1-3	2-7	2-6	5-8	2-6
Unlignified parts		+				+
Druses	+	+	+	+	+	+

U. hyps. = $Urera\ hypselodendron$; Poik. = Poikilospermum; A = adult; J = juvenile; Urticacex = the non-climbing species with unlignified parts in the wood.

Table 4: Comparison of relevant characters of the climbing species of the tribe Urereæ, the genus Poikilospermum, the Cecropiaceæ s. s. (containing the genera Cecropia, Coussapoa, Musanga, Myrianthus and Pourouma) and the Moraceæ (Metcalfe & Chalk, 1950; Topper, pers. comm.) and the Urticaceæ (Metcalfe & Chalk, 1950; Gangadhara & Inamdar, 1977).

Wood	URTICACEÆ	UREREÆ	Poikilosp.	CECR. s.s.	MORACEÆ
Vessel diameter (µm) Vessel member length (µm) Vessels per sq. mm Fibre length (µm) Fibre pit location Ray type (Kribs, 1935) Rays per mm Unlignified parts Druses	100-200 300-500 1-6(15) 750-1 500 R, T He I, Ho 1-3 +/ +/	290-350 375 4-10 630-810 R, T II Ho II 1-3 + +	260-310 325-385 5-9 550-870 R, T Ho II 1-3 +	120-310 400-850 1-6 940-2100 R He II 3-9	100-200 300-500 1-5 600-1900 R He I, II 3-11
Leaf					
Stomatal type Bundle sheath extensions Long shaped cystoliths Raphides	ani-ano +/ +/	ani (ano) ++++	ani +/	ano +	ano (ani) +

Stomatal types: ano = anomocytic; ani = anisocytic (Metcalfe & Chalk, 1979).

II. LEAF ANATOMICAL DESCRIPTIONS

1. Cecropia Loefling - Pl. 4, 2, 3.

Studied: 4 species, 5 specimens.

In surface view: Indumentum of thin, frizzed, unicellular, arachnoid hairs, abaxial; unicellular, needle-shaped, rarely hooked hairs (mainly on abaxial surface, rarely also on adaxial surface); adaxial, glandular hairs with multicellular, globular heads on 3-5-celled, uniseriate stalks, solitary or in groups of 2-4; abaxial, uniseriate 5-8-celled, curved, glandular hairs with or without globular to elongated heads; and mostly abundant, conical papillæ containing lithocysts. Müllerian bodies and pearl glands present or absent. Epidermal cells polygonal; adaxial cells overlying large crystalliferous mesophyll cells forming a rosette. Stomata almost entirely confined to abaxial surface, anomocytic, average length of guard cell pairs 15-20 µm, average width 12-18 µm. Hydathodes formed by 10-15 water pores each, present or absent on adaxial surface. Minor veins usually very prominent in abaxial epidermis.

In transverse section: Lamina bifacial. Epidermal cells small, especially abaxially between the veins. Adaxial epidermal cells sometimes with periclinal division walls and/or mucilaginous inner walls. Stomata raised above level of unspecialized cells. Adaxial hypodermis of 1 or 2 layers of parenchyma cells, including mucilage cells present or absent. Mesophyll consisting of one layer of palisade cells (sometimes subdivided), compact spongy tissue, with or without an intermediate layer in between. Veins with sclerenchymatous vertical bundle sheath extensions (touching adaxial hypodermis and abaxial epidermis). Midrib with a flat or raised adaxial surface and a prominently raised abaxial surface; peripheral ground tissue parenchymatous to collenchymatous, interspersed with mucilage cells; vascular system composed of a closed or variously interrupted cylinder, partly or wholly surrounded by sclerenchyma fibres, and enclosing a parenchymatous "pith". Vascular system of petiole similar. Crystals present as large druses throughout mesophyll and as small to large druses in petiole and midrib.

Note: Müllerian bodies (Schimper, 1888; Rickson, 1971, 1976) are ovoid or pear-shaped, 3 × 1 mm, deciduous, multicellular, food bodies. The apex of each body communicates with the exterior via a stoma (Metcalfe & Chalk, 1950). The bodies on the lower surface of the base of the petiole are situated amongst a velvety covering of uniseriate hairs (Baily, 1922; Janzen, 1973). Sometimes these trichilia are reduced or even lacking (Berg, 1980, 1981; Burger, 1977).

Pearl glands (Meyen, 1837) are trichomes on the petiole and the blade. They are non-secretory, large, vacuolate cells, containing lipid droplets and a small number of glycogen

plastids (Rickson, 1976).

2. Coussapoa Aublet

Studied: 11 species, 18 specimens.

In surface view: Indument of thin, frizzed, unicellular arachnoid hairs, abaxially present or absent; unicellular needle-shaped, often wavy hairs (mostly on abaxial surface, rarely also on adaxial surface); adaxial, glandular hairs with multicellular, globular heads on 3-5-celled, uniseriate stalks, mostly in groups of 2-4, present or absent; abaxial, uniseriate, 6-10-celled, curved, glandular hairs with globular to elongated heads, mostly present; and conical papillæ sometimes present. Epidermal cells polygonal; adaxial cells overlying large crystalliferous mesophyll cells forming a rosette. Stomata confined to abaxial surface, anomocytic, average length of guard cell pairs 15-20 µm, average width 12-18 µm. Hydathodes formed by 10-15 water pores each, present or absent on adaxial surface. Minor veins usually very prominent in abaxial epidermis.

In transverse section: Lamina bifacial. Epidermal cells small, especially abaxially between the veins. Adaxial epidermal cells sometimes with silicified outer walls. Stomata sometimes raised above level of unspecialized cells. Adaxial hypodermis of 2 or 3 layers of parenchyma cells, including mucilage cells except in C. villosa. Mesophyll consisting of one layer of palisade cells (sometimes subdivided), loose spongy tissue, with or without an intermediate layer in between. Veins with sclerenchymatous vertical bundle sheath extensions (touching adaxial hypodermis and abaxial epidermis). Midrib with a flat or raised adaxial surface and a prominently raised abaxial surface; peripheral ground tissue parenchymatous to collenchymatous, interspersed with mucilage cells; vascular system composed of a closed or variously interrupted cylinder, partly or wholly surrounded by sclerenchyma fibres, and enclosing 1 or 2 rows of bundles which are situated in the same direction as the most abaxial bundle of the cylinder (Pl. 1, 3). Vascular system of petiole similar. Crystals present as druses throughout the mesophyll, in petiole and midrib; rhombic crystals sometimes present.

3. Musanga R. Brown

Studied: 1 species, 2 specimens.

In surface view: Indumentum of thin, frizzed, unicellular arachnoid hairs, abaxial; unicellular needle-shaped hairs on abaxial surface; adaxial, glandular hairs with multicellular, globular heads on 3-5-celled, uniseriate stalks, in groups of 2-7; adaxial, uniseriate, 5-8-celled, curved, glandular hairs with or without globular to elongated heads; and conical papillæ. Epidermal cells polygonal; stomata entirely confined to abaxial surface, anomocytic, average length of guard cell pairs 15-20 μm, average width 12-18 μm. Minor veins usually very prominent in abaxial epidermis.

In transverse section: Lamina bifacial. Epidermal cells small. Adaxial epidermal cells sometimes with silicified outer walls. Adaxial hypodermis of 2 layers of parenchyma

cells, including mucilage cells. Mesophyll consisting of one layer of palisade cells, compact spongy tissue, with or without an intermediate layer in between. Veins with parenchymatous to collenchymatous vertical bundle sheath extensions (touching adaxial hypodermis and abaxial epidermis). Midrib with grooved adaxial surface and prominently raised abaxial surface; peripheral ground tissue parenchymatous to collenchymatous, interspersed with mucilage cells; vascular system composed of a closed or variously interrupted cylinder, partly or wholly surrounded by sclerenchyma fibres, and enclosing a parenchymatous "pith". Vascular system of petiole similar. Crystals present as druses throughout mesophyll, in petiole and midrib; rhombic crystals sometimes present in midrib.

4. Myrianthus Pal. Beauv. — Pl. 4, 1.

Studied: 3 species, 3 specimens.

In surface view: Indumentum of thin, frizzed, unicellular arachnoid hairs, abaxial; unicellular, sometimes bicellular, needle-shaped hairs (mainly on abaxial surface, rarely also on adaxial surface); adaxial, glandular hairs with multicellular, globular heads on 3-5-celled, uniseriate stalks, in groups of 2-7; abaxial, uniseriate, 5-8 celled, curved, glandular hairs with or without globular to elongated heads. Epidermal cells polygonal; abaxial cells partly papillated. Stomata entirely confined to abaxial surface, anomocytic, average length of guard cell pairs 15-20 µm, average width 12-18 µm. Minor veins usually very prominent in abaxial epidermis.

In transverse section: Lamina bifacial. Epidermal cells small, especially abaxially between the veins. Adaxial hypodermis of 1 or 2 layers of parenchyma cells. Mesophyll consisting of one layer of palisade cells, compact spongy tissue, with or without an intermediate layer in between. Veins with sclerenchymous vertical bundle sheath extensions (touching adaxial hypodermis and abaxial epidermis). Midrib with a flat adaxial surface and prominently raised abaxial surface; peripheral ground tissue parenchymatous to collenchymatous, interspersed with mucilage cells; vascular system composed of a closed or variously interrupted cylinder, partly or wholly surrounded by sclerenchyma fibres, and enclosing 1 or 2 rows of bundles which are situated in the same direction as the most abaxial bundle of the cylinder (Pl. 1, 3). Vascular system of petiole composed of a closed cylinder. Crystals present as large druses throughout the mesophyll and as small to large druses and rhombic crystals in petiole and midrib.

5. Poikilospermum Zipp. ex Miquel — Pl. 3, 3, 4.

Studied: 4 species, 5 specimens.

In surface view: Indumentum of unicellular, needle-shaped hairs sometimes on abaxial surface; abaxial and adaxial, glandular hairs with unicellular or uniseriate heads (shape varies in the different species) on unicellular stalks. Epidermal cells polygonal. Stomata confined to the abaxial side, anisocytic, average length of guard cell pairs 25-30 µm,

average width 20-25 µm. Hydathodes formed by 20-40 water pores diffusely distributed on adaxial surface.

In transverse view: Lamina bifacial. Epidermal cells tall. Abaxial epidermal cells sometimes with periclinal division walls. Adaxial hypodermis of 1, 2 or 3 layers of parenchyma cells, including mucilage cells. Mesophyll consisting of two layers of palisade cells, and loose spongy tissue. Midrib with a flat adaxial surface and a prominently raised abaxial surface; peripheral ground tissue parenchymatous to collenchymatous, interspersed with mucilage cells; vascular system composed of one or more parallel arcs, partly or wholly surrounded by sclerenchyma fibres; the upper arc with strongly incurved edges. Vascular system of petiole composed of one or more closed or variously interrupted parallel arcs. Crystals present as numerous druses throughout mesophyll, in petiole and midrib; raphides sometimes present in mesophyll. Cystoliths on both surfaces, on adaxial surface arranged pointing towards hydathodes (Pl. 3,4) and sometimes penetrating deeply into mesophyll, on abaxial surface along midrib and veins; in shape punctiform, elongate or stellate.

6. Pourouma Aublet - Pl. 4, 4.

Studied: 10 species, 12 specimens.

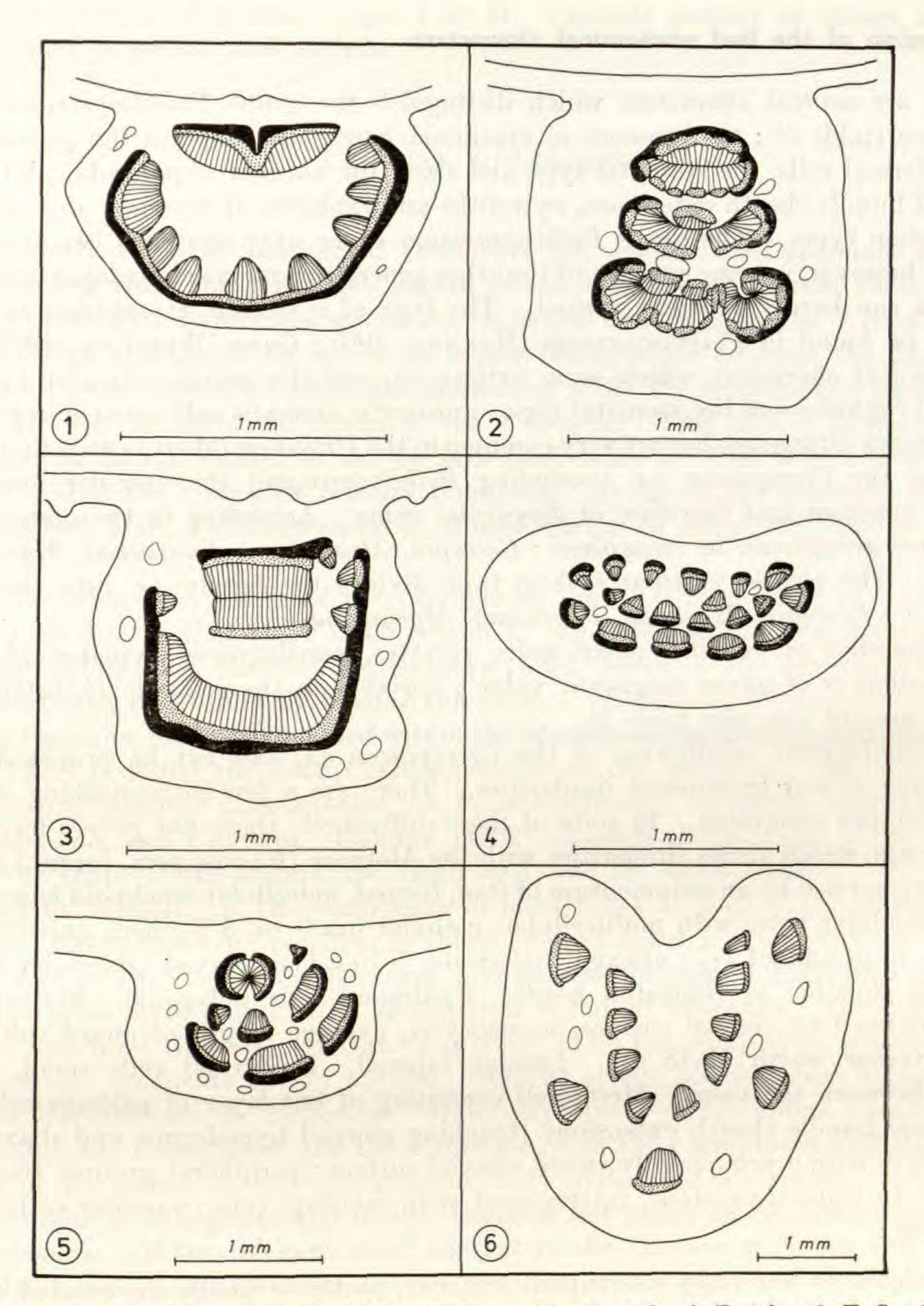
In surface view: Indumentum of thin, frizzed, unicellular arachnoid hairs, abaxial; unicellular needle-shaped hairs (on abaxial surface and sometimes on adaxial surface); adaxial, glandular hairs with multicellular, globular heads on 3-7-celled, uniseriate stalks, in groups of 2-7, often in pits; abaxial, uniseriate, 5-8-celled, curved, glandular hairs with or without globular to elongated heads; and conical papillæ, sometimes present. Epidermal cells polygonal; abaxially rarely papillated; adaxial cells overlying large crystalliferous mesophyll cells forming a rosette. Stomata entirely confined to abaxial surface, anomocytic, average length of guard cell pairs 15-20 µm, average width 12-18 µm. Hydathodes formed by 10-15 water pores each, sometimes present on adaxial surface. Minor veins very prominent in abaxial epidermis.

In transverse section: Lamina bifacial. Epidermal cells small, especially between the veins. Stomata raised on pedestals. Adaxial hypodermis of 1 or 2 layers of parenchyma cells. Mesophyll consisting of one layer of palisade cells, loose or compact spongy tissue, with or without an intermediate layer in between. Veins with sclerenchymatous vertical bundle sheath extensions (touching adaxial hypodermis and abaxial epidermis). Midrib with a flat or grooved adaxial surface and a prominently raised abaxial surface; peripheral ground tissue parenchymatous to collenchymatous, interspersed with mucilage cells; vascular system composed of 2 or 3 closed or variously interrupted flattened cylinders, partly or wholly surrounded by sclerenchyma fibres (Pl. 1, 2). Vascular system of petiole composed of a closed or variously interrupted cylinder, partly or wholly surrounded by sclerenchyma fibres, and enclosing a parenchyma "pith". Crystals present as large druses throughout mesophyll; sometimes druses in midrib, petiole and/or adaxial epidermis.

Table 5: Comparison of some leaf anatomical characters of the Cecropiaceæ s.l.

	Cecropia	Coussapoa	Musanga	Myrian thus	Pourouma	Poikilos per mum
INDUMENTUM:						
arachnoid hairs needle-shaped hairs conical papillae similar glands on both		(+) (+)			+ + (+)	(+)
surfaces						+
EPIDERMIS:						
cell size silicified outer walls rosettes	small (+) (+)	small (+) (+)	small (+)	small —	small +	large —
STOMATA:						
type size (µm) raised Hydathodes	anomocytic 15-20 (+) (+)	anomocytic 15-20 (+) (+)	anomocytic 15-20	anomocytic 15-20	anomocytic 15-20 + (十)	anisocytic 25-30 +
Hypodermis:						
number of cell layers mucilage cells	0-2	2-3	2	1-2	1-2	1-3
MESOPHYLL:						
number of palisade cell layers intermediate layer spongy tissue type Bundle sheath extensions Midrib vascular system ty Petiole vasc. system type Crystal type Cystoliths in lamina		$\begin{array}{c} 1 \\ (+) \\ \text{loose} \\ + \\ \text{II} \\ \text{II} \\ \text{dr/rh} \end{array}$	$\begin{array}{c} 1\\ -\\ \text{compact}\\ +\\ I\\ I\\ \text{dr/rh} \end{array}$	1 (+) compact + II I dr/rh	1 (+) loose/compact + III I dr	loose plate 1, 5 plate 1, 6 dr/raphides

Legend: dr = druses, rh = rhombic crystals, (+) = absent or present.



Pl. 1. — Cecropia latiloba Miq.: 1, T. S. of the midrib. — Pourouma lævis Benth.: 2, T. S. of the midrib. — Myrianthus serratus (Trec.) B. & H.: 3, T. S. of the midrib. — Coussapoa nitida Miq.: 4, T. S. of the petiole. — Poikilospermum inæquale Chew: 5, T. S. of the midrib; 6, T. S. of the petiole.

Discussion of the leaf anatomical characters

There are several characters which distinguish the genus Poikilospermum from the other genera (table 5): the presence of arachnoid hairs, the shape of the glands, the size of the epidermal cells, the stomatal type and sizes, the number of palisade cell layers, the presence of bundle sheath extensions, cystoliths and raphides (if present), and the vascular bundle system types. The genus Poikilospermum is the only one with two true palisade cell layers, however, in some samples of the other genera the intermediate layer is so palisade-like that it can hardly be distinguished. The type of cystoliths as occuring in the Urticaceæ can be found in Poikilospermum (Renner, 1907; Chew Wee-Lek, 1963). Other leaf anatomical characters which seem urticaceous are the petiole vascular system, the presence of raphides and the stomatal type: anisocytic stomata only occur in a part of the genus Dorstenia (Moraceæ), but are very common in the Urticaceæ (Metcalfe & Chalk, 1950).

Within the Cecropiace s.s. (excluding Poikilospermum) the vascular bundle types are rather constant and therefore of diagnostic value. According to the midrib vascular system three groups can be recognized: Cecropia, Musanga — Coussapoa, Myrianthus — Pourouma. The petiole vascular system type divides the family s.s. into two groups:

Coussapoa — Cecropia, Musanga, Pourouma, Myrianthus.

The presence of silicified outer walls, rosettes, mucilaginous hypodermal cells and conical papillae is of minor diagnostic value: variable on the genus level, totally lacking

or always present (see also table 5).

An unambiguous subdivision of the Cecropiaceæ s.s. can not be proposed, because of the degree of leaf anatomical similarities. There are a few distinguishing characters, but they are not correlated. In spite of these differences, these five genera form a homogeneous group, which shows similarities with the Moraceæ (Kloos, pers. comm.) and which can be characterized by an indumentum of thin, frizzed, unicellular arachnoid hairs, abaxial; adaxial, glandular hairs with multicellular, globular heads on 3-7-celled, uniseriate stalks, solitary or in groups of 2-7; abaxial, uniseriate, 5-10-celled, curved, glandular hairs with or without globular to elongated heads. Epidermal cells polygonal. Stomata almost entirely confined to abaxial surface, anomocytic, average length of guard cell pairs 15-20 µm, average width 12-18 µm. Lamina bifacial. Epidermal cells small, especially abaxially between the veins. Mesophyll consisting of one layer of palisade cells. Veins with vertical bundle sheath extensions (touching adaxial hypodermis and abaxial epidermis). Midrib with a prominently raised abaxial surface; peripheral ground tissue parenchymatous to collenchymatous, interspersed with mucilage cells; vascular system divided into 3 types:

I: a closed or variously interrupted cylinder, partly or wholly surrounded by sclerenchyma fibres, and enclosing a parenchymatous "pith" (Pl. 1, 1).

II: a closed or variously interrupted cylinder, partly or wholly surrounded by sclerenchyma fibres, and enclosing 1 or 2 rows of bundles which are situated in the same direction as the most abaxial bundle of the cylinder (Pl. 1, 3).

III: 2 or 3 closed or variously interrupted flattened cylinders, partly or wholly surrounded by sclerenchyma fibres (Pl. 1, 2).

Vascular system of petiole: type I or II. Crystals present as druses throughout mesophyll, and in petiole and midrib.

GENERAL DISCUSSION

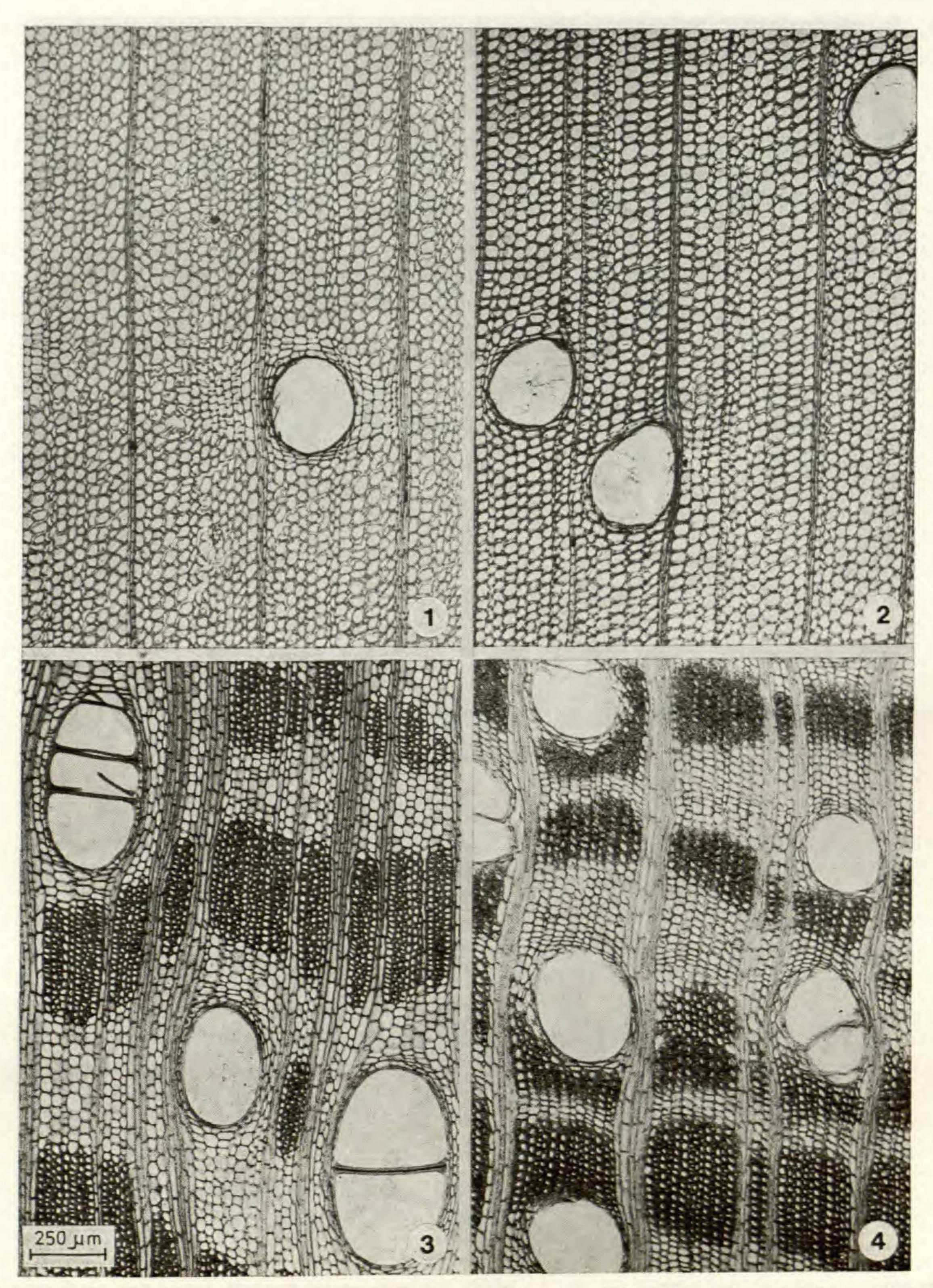
Using wood and leaf anatomical characters the genus Poikilospermum deviates in many characters from the other five genera of the Cecropiaceæ. From these data it is quite clear that Poikilospermum should be classified elsewhere in the Urticales. Chew Wee-Lek (1963) considered the genus as rather intermediate between the Moraceæ and the Urticaceæ: the vegetative parts are moraceous while the reproductive parts are urticaceous. He has classified the genus in the Urticaceæ. This idea is supported by the results obtained in this study, as can be seen from table 4. The proper place of Poikilospermum within the Urticaceæ remains doubtfull. As far as the anatomy of the Urticaceæ is studied here, Poikilospermum is almost similar in wood structure to the genus Urera.

The remaining five genera of the Cecropiaceæ are so homogeneous in wood and leaf anatomical characters that there is no reason to separate one of these genera from this group. The subdivision discribed by Renner (1907) based on the adaxial glands, is not at all supported by the results obtained in this study. Should this wood and leaf anatomical rather homogeneous group be included within the Moraceæ or should it be given family rank? Engler (1889) included the tribe Conocephaloideæ in the Moraceæ. The tribe consisted of the following genera: Conocephalus (= Poikilospermum), Musanga, Myrianthus, Coussapoa, Pourouma, and Cecropia. Corner (1962) transferred this tribe to the Urticaceæ, based on the shape of the stigma, the small seed, and the small embryo. One character not studied by himself, i.e. the occurrence of latex-tubes at least in the primary bark, is moraceous. Latex-tubes are common in the secondary wood of most genera of the Moraceæ but scarce in a few species of Cecropia, and possibly present in Pourouma and Coussapoa. Latex-tubes were found to be absent from wood of Musanga, Myrianthus, Poikilospermum, and the Urticaceæ. In Berg's opinion (1978) the group of six genera (including Poikilospermum) constitute a very natural, coherent group, which merits a rank equal to that of the Moraceæ and the Urticaceæ. However, almost all characters discussed by him occur in either the Cecropiaceæ and the Urticaceæ or in the Cecropiaceæ and the Moraceæ. The group of five genera is very homogeneous in its anatomical characters. Bundle sheath extensions occur in this group and in the Moraceæ but never in the Urticaceæ (including Poikilospermum). Of the relevant wood characters the absence of druses and unlignified parts, the number of rays per mm, the composition of the rays, and the location of the fibre pits restricted to the radial walls, are shared by the Cecropiaceæ s.s. and the Moraceæ. There are neither leaf nor wood anatomical characters that occur exclusively in the Urticaceæ and Cecropiaceæ, and not in the Moraceæ. These results are supported by the students of the wood and the leaf anatomy of the family of the Moraceæ, S. M. C. Topper and A. Kloos (pers. comm.) respectively. As the Cecropiaceæ s.s. constitute a very homogeneous taxon and taking into account the fact that it has almost always been considered a natural group by taxonomists (with the exception of Снеw Wee-Lek, 1963, see intro-

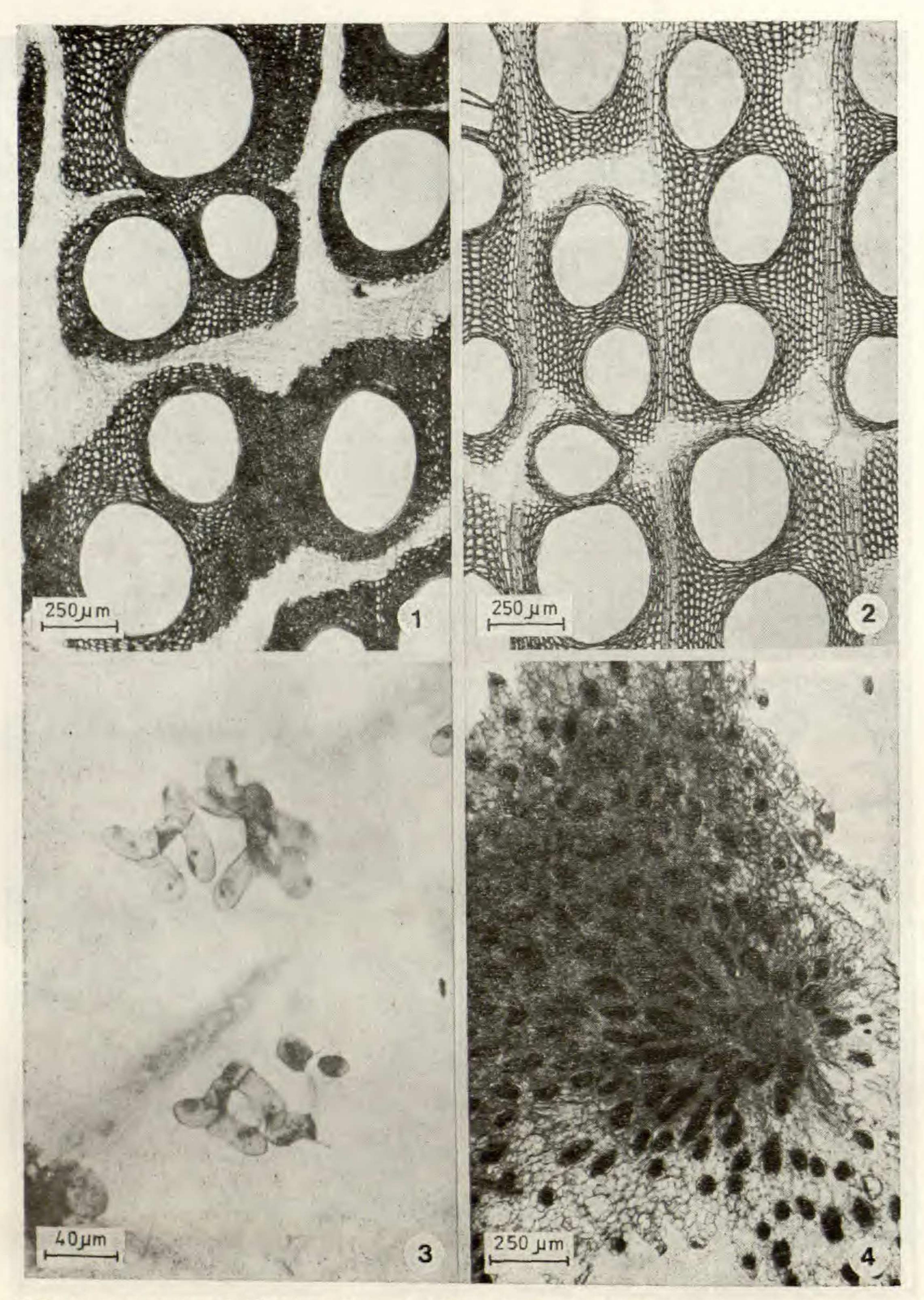
Table 6: Some wood anatomical characters of the Cecropiaceæ.

	con .	SO		T			Y	8	28				es
	vessels	vessels	diam.	member m)	(mm)	(mm)	ray	ray	rays				samples
	number of ve per cluster	number of ve per sq. mm	g. pore d (μm)	3	diam.	fibre length	multiseriate height (µm)	multiseriate width (cells)	ber of r mm	par. distr.	erystal type	tal type	er of san per species
	numb	numb per s	tang.	vessel	fibre	fibre	multis	multis	number per m	tr. pa	crys	crystal	nber per
	Y.	Ĭ.	Y.	LV.	Y.	7	Y.	AV.	Lv.	Pare	Ray	Par.	Nur
CECROPIA													
C. burriada	3(4)	3	175	630	28	1 520	1 050	4	7	v, a			1
C. concolor	2(4)	3	155	505	50	1 100	890	3	7	v	_		1
C. garcia	3(5)	2 3	245 170	500	35	2 000	1 530	3 4	4 5	v, a	- h	rh	1
C. obtusa C. peltata	$3(5) \\ 3(5)$	2	200	10.00	30	The second secon	1 010	3	5	v, a v, a	rh rh	rh	1
C. riparia	2(5)	1	185	570	45	1 660	2 100	3	5	v, a	_	rh	1
C. sciadophylla	3(5)	2	260	620	30	1 780	1 380	4	4	v, a, c	rh	rh	10
C. telealba	3(4)	2	255	515	33	1 500	1 600	4	4	v, a		rh	1
COUSSAPOA													
C. angustifolia	2	3	295	565	25	1 800	960	3	4	a, c	_	rh	1
C. asperifolia	2	2	290	595	25	1 480	1 050	5	6	c, b		rh	1
C. batavorum C. contorta	2	2 5	275 220	520 320	25 20	1 675	1 100 665	4	7	c, b		rh	1
C. comoria C. latifolia	2	2	300	475	20	1 760	1 080	5	5	c, b		rh	1
C. trinervia	2	1	250	550	20	1 370	1 080	4	7	b		rh	1
C. villosa	2	2	220	530		1 600	770	5	6	b	-	rh	1
MUSANGA													
M. cecropioides	2(4)	1	265	550	45	1 500	1 000	3	4	v	rh	rh	9
M. leo-erreræ	2(3)	2	210	460	The second second		920	4	3	v	rh	rh	1
MYRIANTHUS													
M. arboreus	3	3	225	500	25	1 550	1 350	7	6	b	rh	rh	5
M. holstii	3(5)	5	150	420		1 400		6	6	b		rh	3
M. libericus	3(5)	3	200	500					5	b	rh	rh	4
M. serratus	3(5)	4	140	450	25	1 110	1 050	5	5	b	_	rh	1
POIKILOSPERMI	UM												
P. nauclei florum	2	5	260	325		660	COLUMN TO SERVICE AND ADDRESS OF THE PARTY O		2	v, unlign.		rh, dr	1
P. suaveolens	2	9	310	350	25	865	comp.	. 6	2	v, unlign.	-	rh, dr	2
POUROUMA													
P. acuminata	3(4)	5	170	615	1				6	a, c	_	rh	1
P. acutiflora D. aniculata	2(4)	3	195	530					8	a, c	rh	rh	1
P. apiculata P. aspera	2(3)	3	250	580					4	v, a	-		1
P. aspera P. cecropiifolia	2(4)	3	180	650					5	a, c	rh	rh	2
P. chocoana	2	6	175	475					6	a, c	rh	rh rh	1
P. digitata	2(3)	2	190	500	25					a, c	rh	rh	2
P. guianensis	2-3	4	200	680	30	1 300	850	3	5	a, c		rh	4
P. hirsutipetrolata		to the second	220	845			905	3		v, a	rh	rh	1
P. lævis	2(3)	2	240					1		v		rh	1
P. maroniensis P. melinonii	2(3) $2(4)$	3	210	550 620					6	v, a	_	rh	2
P. mellis P. mollis	2(4)		210	620						v, a	_	rh	4
P. mouis P. ovata	2(3)		225	550					5	v, a	-		4
P. substrigosa	2	3	215	600					5	D		rh	1
P. triloba	2(3)		255			The second second	A PARTY		4	a, c	rh	rh	1
							- 200		-	a, c	rh	TH	

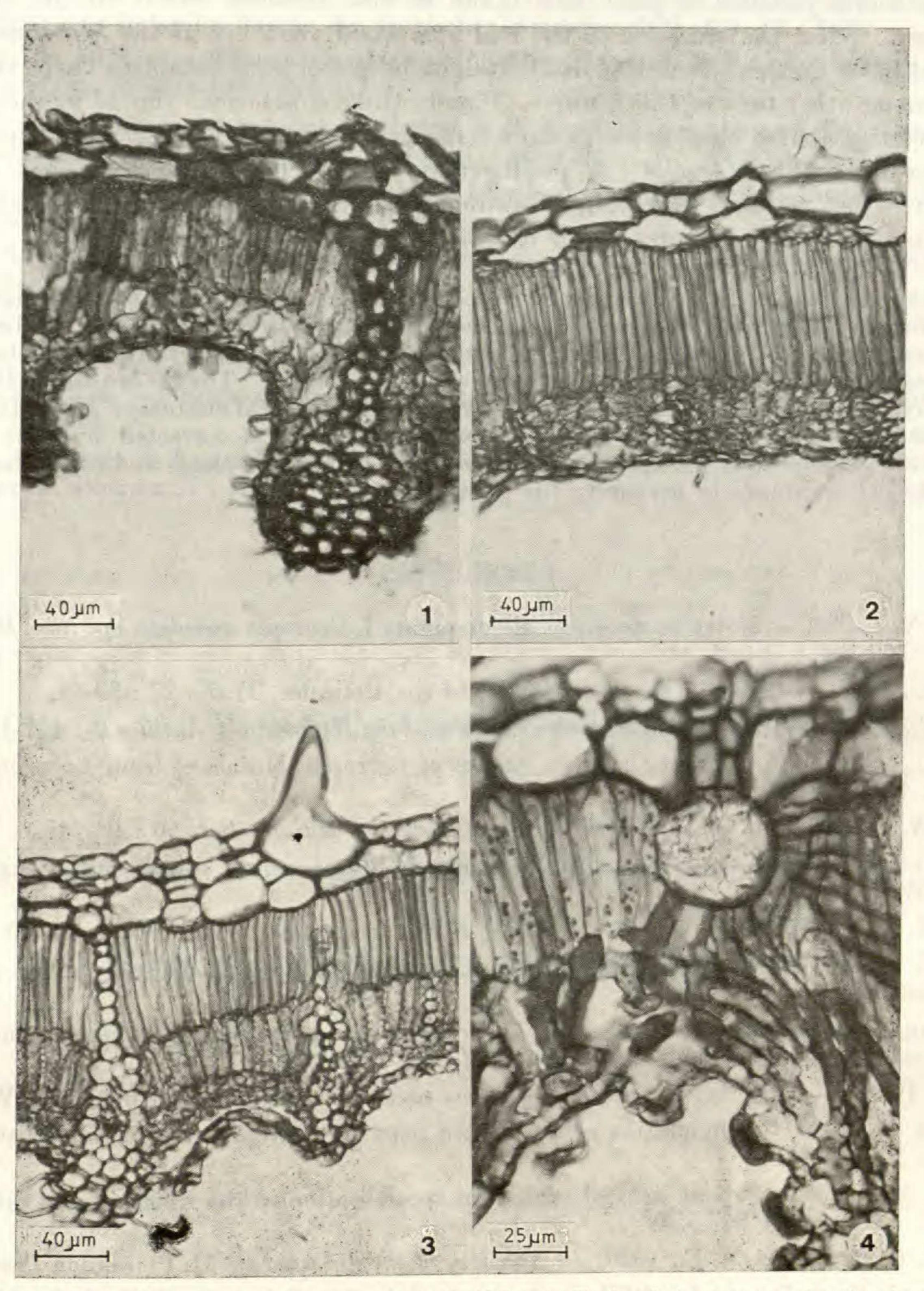
Legend: v = vasicentric; a = aliform; c = confluent; b = banded; unlign. = unlignified apotracheal parer chyma; rh = rhombic crystals; dr = druses; comp. = vertically compounded.



Pl. 2. — Cecropia sciadophylla Mart. (Maguire & al. 55577): 1, T. S. of the wood. — Musanga cecropioides R. Br. (ex MAD-SJRw 15799): 2, T. S. of the wood. — Coussapoa latifolia Aubl. (Lindeman & Heyde 78): 3, T. S. of the wood. — Myrianthus libericus Rendle (ex MADw 36781): 4, T. S. of the wood.



Pl. 3. — Poikilospermum naucleiflorum Euse (RTIw 1507/i22-H 1868-274): 1, T. S. of the wood. — Urera hypselodendron (Horhst.) Wedd. (Schlieben 1721): 2, T. S. of the wood. — Poikilospermum subtrinervium (Miq.) Chew (Mandi 25): 3, glands on adaxial leaf surface. — Poikilospermum amboinense Zipp. ex Miq. (van Leeuwen s.n.): 4, Cystoliths around a hydathode on adaxial leaf surface.



Pl. 4. — Myrianthus arboreus Pal. Beauv. (Deistel 428): 1, T. S. of the lamina showing bundle sheath extension, promine at vein and papillated abaxial epidermal cells. — Cecropia surinamensis Miq. (L.B.B. 12739): 2, T. S. of the lamina showing epidermal cells with silicified outer walls. — Cecropia latiloba Miq. (Pranze & Berg P 17586): 3, T. S. of the lamina showing conical papilla, intermediate layer and some arachnoid hairs. — Pourouma lævis Benth. (Boschwezen 4010): 4, T. S. of the lamina showing a large druse.

duction), it seems justified to place this taxon in the Moraceæ based on the characters studied here. After the students of the leaf and wood anatomy of the Moraceæ, mentioned before, have finished their research it might be possible to establish their taxonomic position among other taxa of this family. Finally these conclusions should be incorporated in a re-investigation of the (flower) characteristics used by Berg (1978) to separate the Cecropiaceæ from the Moraceæ. A phylogenetic approach of this taxon has not been carried out. This requires a profound anatomical investigation of all related taxa, which will be done by Topper (in prep.) and Kloos (in prep.).

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